

1 **Uncovering the hidden niche: incorporating microclimate temperature into species distribution**
2 **models**

3 **Stef Haesen^{1,2}, Jonathan Lenoir³, Eva Gril³, Pieter De Frenne⁴, Jonas J. Lembrechts⁵, Martin**
4 **Kopecký^{6,7}, Martin Macek⁶, Matěj Man^{6,8}, Jan Wild^{6,9}, Koenraad Van Meerbeek^{1,2}**

5 **Corresponding author, OrcID = <https://orcid.org/0000-0002-4491-4213>, stef.haesen@kuleuven.be,*
6 *+32 16 32 24 67*

7 *¹Department of Earth and Environmental Sciences, Celestijnenlaan 200E, 3001 Leuven, Belgium; ²KU*
8 *Leuven Plant Institute, KU Leuven, Leuven, Belgium; ³UMR CNRS 7058 "Ecologie et Dynamique des*
9 *Systèmes Anthropisés" (EDYSAN), Université de Picardie Jules Verne, Amiens, France; ⁴Forest & Nature*
10 *Lab, Department of Environment, Ghent University, Geraardsbergsesteenweg 267, 9090 Melle-*
11 *Gontrode, Belgium; ⁵Research Group PLECO (Plants and Ecosystems), University of Antwerp, 2610*
12 *Wilrijk, Belgium; ⁶Institute of Botany of the Czech Academy of Sciences, Zámek 1, CZ-25243, Průhonice,*
13 *Czech Republic; ⁷Faculty of Forestry and Wood Sciences, Czech University of Life Sciences Prague,*
14 *Kamýcká 129, CZ-165 21, Prague 6 - Suchdol, Czech Republic; ⁸ Department of Botany, Faculty of*
15 *Science, Charles University, Benátská 2, CZ-128 01 Prague 2, Czech Republic; ⁹ Faculty of Environmental*
16 *Sciences, Czech University of Life Sciences Prague, Kamýcká 129, CZ, 165 21 Prague 6 - Suchdol, Czech*
17 *Republic;*

18 **OrcIDs**

19 *Jonathan Lenoir: <https://orcid.org/0000-0003-0638-9582>*
20 *Eva Gril: <https://orcid.org/0000-0002-7340-8264>*
21 *Pieter De Frenne: <https://orcid.org/0000-0002-8613-0943>*
22 *Jonas J. Lembrechts: <https://orcid.org/0000-0002-1933-0750>*
23 *Martin Kopecký: <https://orcid.org/0000-0002-1018-9316>*
24 *Martin Macek: <https://orcid.org/0000-0002-5609-5921>*
25 *Matěj Man: <https://orcid.org/0000-0002-4557-8768>*
26 *Jan Wild: <https://orcid.org/0000-0003-3007-4070>*
27 *Koenraad Van Meerbeek: <https://orcid.org/0000-0002-9260-3815>*

28 **ABSTRACT**

29 Species' environmental niches are conventionally modelled using coarse-grained macroclimate data.
30 These data are known to deviate substantially from local, near-ground and proximal conditions (i.e.,
31 the microclimate), especially so below forest canopies. Here, we aimed to assess the impact of using
32 gridded microclimate data instead of gridded macroclimate data on the performance of species
33 distribution models (SDMs), as well as on the predicted geographical distribution and the derived
34 species response curves of 140 forest specialist plant species across Europe over the 2000-2020
35 period. We performed a comparative study between SDMs constructed with different sets of
36 bioclimatic predictors to separately test the effect of using (i) proximal climate data instead of
37 conventional macroclimatic data and (ii) high-resolution proximal climate data rather than coarse-
38 gridded macroclimatic data. Therefore, we challenged SDMs with: (1) a macroclimatic dataset at a
39 spatial resolution of 1 km × 1 km; (2) an aggregated microclimatic dataset matching the same
40 resolution of 1 km × 1 km; and (3) a microclimatic dataset at a much finer spatial resolution of 25 m ×
41 25 m. We found significant differences in model performance, indicating that microclimate-based
42 SDMs outperform both their macroclimatic and aggregated counterparts. Most importantly, this study
43 makes clear that macroclimate-based SDMs tend to introduce a systematic bias into the perceived
44 species response curves. Additionally, macroclimatic data is unable to identify warm and cold refugia
45 beyond the range edges of species' distributions. We thus conclude that microclimate-based SDMs
46 are a crucial tool to gain peculiar insights regarding biodiversity conservation, which is needed to align
47 management actions and prioritize conservation efforts.

48 **Keywords:** climate change, species distribution modelling, MaxEnt, microclimate, ForestTemp, forest
49 plant species, species response curves, understory temperatures

50 INTRODUCTION

51 Over the last decades, species distribution models (SDMs, also known as ecological niche models or
52 habitat suitability models), have emerged as a central method to project the effects of changing
53 environmental conditions on species' distributions in space and time (Elith & Leathwick, 2009; Guisan
54 & Zimmermann, 2000; Zimmermann et al., 2010). Species distribution models are employed for a wide
55 range of applications that are vital to support conservation decision making (Baker et al., 2021),
56 ranging from quantifying the effects of contemporary climate change on biodiversity (Araújo et al.,
57 2011; Pearce-Higgins et al., 2017) to the management of invasive species (Roy-Dufresne et al., 2019;
58 Srivastava, 2019) and rewilding practices (Jarvie & Svenning, 2018).

59 SDMs commonly are correlative models that infer relationships between species occurrences
60 and the environment using statistical or machine learning methods (Elith & Leathwick, 2009).
61 Conventional SDM practices involve the incorporation of a standard set of bioclimatic variables with
62 a maximal spatial resolution of 30 arc seconds ($\pm 1 \text{ km}^2$ at the equator) such as WorldClim (Fick &
63 Hijmans, 2017; 1 km^2), CHELSA (Karger et al., 2017; 1 km^2) or TerraClimate (Abatzoglou et al., 2018;
64 16 km^2). However, these climatological data are derived from standardized meteorological stations at
65 approximately 2 meters height above short vegetation, exposed to wind, and well away from trees
66 and buildings to minimize any noise generated by microclimatic effects (Jarraud, 2008). Gridded
67 macroclimatic data interpolate such weather stations' data and thus represent the free-air
68 temperature conditions in open ecosystems. Although these data are sufficient to adequately capture
69 changes in free-air temperatures, problems arise when using these data to model the response of
70 species that live close to the ground in topographically heterogenous terrain and/or in ecosystems
71 with trees and shrubs. For instance, within-pixel (1 km^2) variability of mean annual temperatures can
72 be as high as $6 \text{ }^\circ\text{C}$ in mountainous areas, and this might even increase when different land-use types
73 are present within a single grid cell (i.e., forested and non-forested areas; Lenoir et al., 2013). This high
74 thermal variability results from physical processes such as air flow and incoming solar radiation that
75 interact with topographic factors such as slope aspect and surface roughness (Geiger, 1950).
76 Additionally, vegetation cover is known to affect local microclimate temperature (De Frenne et al.,
77 2019; Lenoir et al., 2017). Indeed, it is currently well acknowledged that forests harbour distinct
78 microclimatic conditions owing to the structural complexity of the canopy, resulting in shading and
79 evapotranspirative cooling (Geiger, 1950). Consequently, forest canopies are characterized by their
80 buffering capacities of extreme temperatures in comparison to weather station data, with cooler sub-
81 canopy maximum temperatures and warmer sub-canopy minimum temperatures (De Frenne et al.,
82 2019). In European forests, this difference can add up to 9°C for mean monthly temperatures (Haesen
83 et al., 2021).

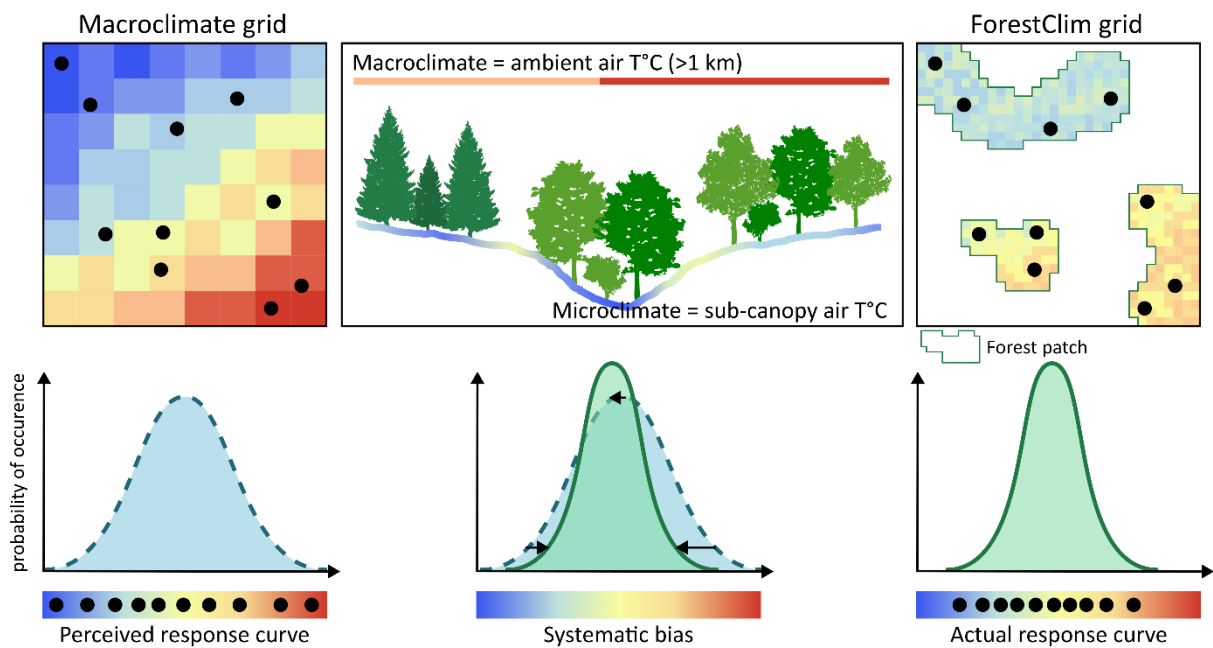
84 It is clear that fine-scale microclimatic data should urgently be used within SDMs – and
85 ecological research in general – as ignoring the mismatch between conventionally-used macroclimatic
86 data and the apparent microclimatic conditions might lead to erroneous predictions, wrong ecological
87 interpretations and, ultimately, questionable conservation decisions (Körner & Hiltbrunner, 2018).
88 Especially under contemporary climate change, where species are shifting their distributions in
89 accordance with the moving isotherms, accurate estimations of species distributions are needed. The
90 correct identification of leading and trailing edges of moving species distributions is of particular
91 interest for conservation since they coincide with the formation and disappearance of suitable habitat,
92 respectively (Greiser et al., 2020). Furthermore, recent studies emphasize on the importance of so-
93 called microrefugia, where species can find more stable climatic conditions in which they can persist
94 for a longer amount of time (Finocchiaro et al., 2022; Nadeau et al., 2022). However, current SDM
95 practices are unable to identify these microrefugia as conventional macroclimate data represents the
96 overarching free-air temperatures rather than the local temperatures inside these microrefugia
97 (Lenoir et al., 2017), which act at high-spatial resolution.

98 With recent advances in microclimate modelling (Gril et al., 2023; Maclean, 2019), there has
99 been an increase in the usage of microclimate in SDMs (Graae et al., 2018; Greiser et al., 2020; Lenoir
100 et al., 2017). Nevertheless, current studies have not yet been able to incorporate fine-scale
101 microclimatic data over continental extents (Lembrechts, Nijs, & Lenoir, 2018). However, with the
102 recent advent of sub-canopy microclimate layers for European forests at 25 m × 25 m resolution, a
103 new avenue of species distribution modelling can be explored (Haesen et al., 2023).

104 Here, we compared SDMs constructed with different sets of bioclimatic variables to separately
105 test the effect of using (i) proximal climate data instead of conventional macroclimatic data and (ii)
106 high-resolution proximal climate data rather than coarse-gridded macroclimatic data. We therefore
107 challenge SDMs with (1) a macroclimatic dataset at a spatial resolution of 1 km × 1 km; (2) an
108 aggregated microclimatic dataset matching the resolution of the macroclimatic dataset, yet using
109 proximal below-canopy temperatures; and (3) a microclimatic dataset at a spatial resolution of 25 m
110 × 25 m, matching the resolution of understory vegetation communities and using the proximal below-
111 canopy temperatures. Note that we did not opt to include a high-resolution macroclimatic dataset
112 (i.e., 25 m × 25 m; topographically downscaled) within this comparative study as this would be
113 representative for topoclimatic conditions, which are proximal as well. Additionally, topoclimate
114 would not capture the influence of the canopy cover on below-canopy temperatures.

115 For 198 forest specialist plant species, we aimed to assess the impact of large-scale, gridded
116 microclimate data on the performance of SDMs as well as on their predicted geographical distribution.
117 Furthermore, we compared species thermal response curves constructed with the three methods for

118 the forest specialist plant species, and analysed their behaviour at their range edges. As forests are
 119 known to buffer temperatures, forest specialist plant species respond to warmer minimum
 120 temperatures and lower maximum temperatures as perceived by the free-air temperature data.
 121 Therefore, we hypothesize that (1) the actual thermal response curves of forest specialist species are
 122 narrower than the perceived thermal response curves (Figure 1). Intuitively, the incorrectly modelled
 123 thermal response curve would result in an overestimation of species range distributions when using
 124 macroclimate data. However, the resultant of the effects of the incorrectly modelled species response
 125 and the 'incorrect' macroclimate data on the predicted distribution may lead to deviation from this
 126 expectation. Therefore, our null hypothesis is that (2) the effect of the incorrectly modelled species
 127 response will be cancelled out by the 'incorrect' macroclimate data, so that distributions modelled by
 128 macroclimate and microclimate data will not differ. Finally, assuming that species are constrained by
 129 the maximum temperature at the trailing edge of their distribution and in the minimum temperature
 130 at the leading edge, we hypothesize that (3) populations of forest specialist species survive in local
 131 microrefugia, which are cooler than the surrounding area at the trailing edge and warmer than the
 132 surrounding area at the leading edge.



133

134 *Figure 1: Design of this comparative study, where we compared species distribution models with different set-*
 135 *ups of climatic data. As forests are known to buffer sub-canopy temperatures, forest specialist plant species*
 136 *respond to warmer minimum temperatures and lower maximum temperatures as perceived by the free-air (i.e.*
 137 *macroclimate) temperature data. Therefore, we hypothesize that the actual thermal response curves of forest*
 138 *specialist species are narrower than the perceived thermal response curves. Black points indicate (simulated)*
 139 *species occurrences (adapted from Lenoir et al., 2017).*

140 **METHODS**

141 **Study area & species selection**

142 Our study area encompasses all 27 EU countries, plus Albania, Andorra, Bosnia and Herzegovina,
143 Kosovo, Liechtenstein, Montenegro, North Macedonia, Norway, San Marino, Serbia, Switzerland and
144 the United Kingdom. The Canary Islands and Azores, as well as Europe's overseas territories were
145 excluded from the analysis.

146 Forest specialist species were selected based on the European forest vascular plant species
147 list, which is based on vegetation databases, literature and expert knowledge (Heinken et al., 2022).
148 From this list, we first selected shrub and herb species, which – unlike tree species – usually complete
149 their entire life cycle within the forest understory layer, thus experiencing forest microclimate
150 dynamics (Caron et al., 2021). Subsequently, we selected the species categorized as forest specialists
151 (i.e. categories 1.1 and 1.2 in Heinken et al., 2022) throughout their entire range, meaning that these
152 species occur only in closed-canopy forests, forest edges or forest openings. The final selection
153 encompassed 198 forest specialist species (Table S1).

154 **Environmental predictors**

155 Three different sets of bioclimatic temperature-related variables (i.e., macroclimatic, aggregated
156 microclimatic and microclimatic) were used to construct our SDMs, starting from the conventional set
157 of eleven bioclimatic temperature variables. However, we excluded mean temperature of the wettest
158 quarter (BIO8) and mean temperature of the driest quarter (BIO9) as these were recently criticized for
159 their use within species distribution models (Booth, 2022). As the available CHELSA and WorldClim
160 data are not fully covering our study period (2000-2020), we used TerraClimate to construct the
161 ‘macroclimatic dataset’ at the typical spatial resolution of 1 km² as used in conventional SDMs.
162 TerraClimate bioclimatic variables covering the 2000-2020 period are available at a spatial resolution
163 of 16 km² and thus were spatially downscaled to a spatial resolution of 1 km². To do this, we first
164 calculated, for each 1 km² grid cell, the difference between the bioclimatic variables of TerraClimate
165 (1970-2000 period; 16 km²) and WorldClim (1970-2000 period; 1 km²) and then added these offset
166 values or anomalies to the TerraClimate bioclimatic variables for the 2000-2020 period to come to a
167 final macroclimate layer of 1 km² resolution. This method assumes that the offset values from the
168 long-term average period of 1970-2000 are still valid for the period 2000-2020.

169 The ‘microclimatic dataset’ consists of the original bioclimatic variables provided within
170 ForestClim, a new high-resolution dataset of forest understory temperature for all European forests
171 at a spatial resolution of 25 m × 25 m derived from the ForestTemp model (Haesen et al., 2021, 2023).
172 Briefly, ForestTemp was created by combining more than 1,200 time series of *in situ* near-surface

173 forest temperatures from across Europe with topographical, biological and macroclimatic predictors
174 in a machine learning model. The ‘aggregated dataset’ was generated by aggregating (i.e. averaging)
175 the ForestClim bioclimatic variables to a 1 km² resolution.

176 Each set of bioclimatic temperature variables was complemented with the conventional set
177 of eight bioclimatic precipitation variables. Note that we omitted precipitation of the warmest quarter
178 (BIO18) and precipitation of the coldest quarter (BIO19) for similar reasons discussed by Booth (2022).
179 The six remaining variables were calculated from TerraClimate precipitation data for the 2000-2020
180 period and disaggregated to match the spatial resolution of each bioclimatic set. Finally, edaphic
181 variables were added, since soil data often increase model performance (Hageer et al., 2017). Based
182 on their effects on plant demography, we selected four soil variables: bulk density (bdod; cg/cm³),
183 which reflects the soil porosity; soil clay content (clay; g/kg), which reflects the soil texture; pH H₂O
184 (pH; unitless); and cation exchange capacity (cec; mmol_c/kg; Hageer et al., 2017). The soil raster layers
185 were downloaded from the SoilGrids database (Poggio et al., 2021) at a resolution of 250 m for three
186 different depths: 0-5 cm; 5-15 cm; and 15-30 cm. These three layers were averaged into one single
187 layer representing the depth from 0 cm to 30 cm, with the exception of pH (i.e., a logarithmic scale),
188 which was aggregated using the median value over the three layers.

189 To help reduce overfitting of SDMs, multicollinearity between the predictors was assessed
190 using a pairwise Spearman correlation test (Figure S1). Highly correlated variables (Spearman
191 correlation coefficients > 0.7) were removed from the analysis in order to reach the most
192 parsimonious model (Dormann et al., 2013). When excluding one of the correlated covariate pair, we
193 retained variables which are known to be more important for plant species distribution (Macek,
194 Kopecký, & Wild, 2019) and which are important for our further analyses (e.g., BIO5 & BIO6). The final
195 selection of covariates encompassed two temperature variables (maximum temperature of the
196 warmest month (BIO5) and minimum temperature of the coldest month (BIO6)), two precipitation
197 variables (mean annual precipitation, (BIO12) and precipitation seasonality (BIO15)) and two edaphic
198 variables (cation exchange capacity and soil clay content). All covariate layers were projected in an
199 equal-area projection (epsg:3035; ETRS89/LAEA).

200 **Species occurrence data**

201 Georeferenced occurrence data for 198 forest plant species were downloaded from the Global
202 Biodiversity Information Facility on the 13th of September 2022 (<https://doi.org/10.15468/dl.kf533a>).
203 To improve data quality for each species, the occurrence data were filtered in the following sequential
204 steps: (1) only records of ‘human observations’ were selected; (2) records with an unknown
205 coordinate uncertainty or coordinate uncertainty larger than 25 m (i.e., the pixel size) were excluded;
206 (3) records located at country or capital centroids and biodiversity institutions (e.g., botanical gardens)

207 were omitted (Cheng et al., 2021); (4) duplicate records were removed; (5) records outside our study
208 area were deleted; (6) only records observed during our climatic reference period (2000-2020) were
209 selected; (7) records were spatially thinned to one random observation per 25 m × 25 m grid cell; and
210 (8) we omitted species with less than 50 cleaned occurrence records, which has been postulated as a
211 minimum standard to build robust species distribution models (van Proosdij et al., 2016; Wisz et al.,
212 2008). Finally, we maintained occurrence data for 140 out of 198 species (Table S1). Note that exactly
213 the same occurrence datasets are needed over the different climatic set-ups to have comparable
214 model outputs. Here, we decided to work with occurrence datasets that underwent a cleaning
215 protocol based upon the characteristics of the microclimatic dataset (i.e., maximum coordinate
216 uncertainty of 25 m, and spatial thinning to a 25 m × 25 m grid cell).

217 **Species distribution modelling**

218 We used MaxEnt, a presence-background algorithm that combines species presence-only data with
219 environmental predictors for the current climate to predict the environmental suitability of each study
220 species across our study area (Phillips, et al., 2017; Phillips & Dudík, 2008). We did that for each of the
221 three sets of bioclimatic variables (i.e., the macroclimatic set, the aggregated microclimatic set and
222 the microclimatic set), thus generating three sets of habitat suitability maps for each study species.
223 Background data were generated by sampling an equal amount of background points as occurrence
224 points (i.e., so that species prevalence equals 50%) based on a 2D kernel-density estimate of the
225 occurrence points (Venables & Ripley, 2002), meaning that the spatial density of the background
226 points is proportional to the spatial density of occurrence points for a given species, thereby
227 accounting for spatial bias in the occurrence points (Lake et al., 2020; Vollerling et al., 2019).

228 Although widely-used in scientific research, MaxEnt could suffer from issues like spatial bias
229 and bad model performance (Radosavljevic & Anderson, 2014). To deal with the problem of spatial
230 bias, we conducted spatially independent evaluations in ENMeval2.0 (Kass et al., 2021; Muscarella et
231 al., 2014) using block cross-validation and allocating 80% of our occurrence points to this cross-
232 validation procedure (20% is kept for independent evaluation). Furthermore, model performance was
233 improved by tuning the model settings in ENMeval2.0 rather than working with the default settings
234 of MaxEnt. This was implemented by means of a grid search over the possible values of the two
235 hyperparameters: feature classes (Linear, Quadratic, Product) and regularization multipliers (0.5, 1, 2,
236 3, 4 and 5). Linear, quadratic and product features were selected to allow for linear and quadratic
237 relationships as well as interactions among predictors (Merow, Smith, & Silander, 2013).
238 Regularization multipliers, on the other hand, control model complexity and overfitting. The larger
239 these regularization multipliers, the smoother the model predictions.

240 **Model performance & sensitivity**

241 In order to customize the settings for the feature classes and the regularization multipliers, a total of
242 42 different models were run for every single species. The Akaike Information Criterion for small
243 sample sizes (AIC_c) was used to select the best candidate models (Burnham & Anderson, 2004). Next,
244 model performance was assessed using the Continuous Boyce Index (CBI), instead of the commonly-
245 used area under the receiver-operating characteristic curve (AUC). The latter has recently been shown
246 to be biased in presence-only models and should therefore be avoided (Jiménez & Soberón, 2020).
247 The CBI is a threshold-independent metric that represents the relationship between predicted habitat
248 suitability and the distribution of occurrence records (Hirzel et al., 2006). Additionally, we calculated
249 the sensitivity enabling us to quantify how good our model is able at distinguishing true positives from
250 false negatives. Both were calculated based on the independent 20% subset of the data.

251 Finally, we used Bayesian regression models (BRMs) in order to assess differences in model
252 performance and sensitivity between SDMs constructed using the three different types of climate
253 data. We opted for BRMs as they are able to account for data dependencies (i.e., values clustered
254 within species), unequal variances among groups and skewed distributions. The final model structures
255 are added in Table S2. Both sensitivity and CBI were modelled with a beta distribution. CBI was
256 rescaled between 0 and 1 before analysis. Bayesian regression models were run using the *brms*
257 package (Bürkner, 2021). All models were first run using standard priors and with 2 chains, 10,000
258 iterations and a warm-up of 1000 runs. When models did not converge, the flat priors were replaced
259 by weakly informative priors (Table S2) and the models were run again with 4 chains. The final models
260 converged with \hat{R} values close to 1 (Gelman and Rubin's diagnostic) and all bulk and tail effective
261 sample sizes of the means were greater than 2500. When the highest posterior density intervals ($\alpha =$
262 0.05) of the contrasts, calculated using the *emmeans* package (Lenth, 2021), did not overlap with zero,
263 contrasts are considered 'significant'.

264 **Model predictions**

265 Habitat suitability was predicted for each species and for each of the three sets of bioclimatic
266 temperature variables (macroclimatic, aggregated microclimatic and microclimatic) for the 2000-2020
267 period. Furthermore, we transformed the logistic maps (i.e., probability values for habitat suitability)
268 to binary (presence-absence) maps using the 10% training presence as a threshold, meaning that the
269 suitable area contains 90% of the original occurrence records (Benito, Cayuela, & Albuquerque, 2013).

270 To compare between model predictions from SDMs constructed with different climate
271 sources and resolutions, we calculated both the potential suitable area and the potential latitudinal
272 range of each species. Note that we (i) disaggregated the binary maps derived from macroclimatic and

273 aggregated data (1 km × 1 km) to the finer resolution (25 m × 25 m), and (ii) masked out all non-forest
274 pixels to make a valid comparison between the three climate types. First, the potential suitable area
275 (km²) was calculated as the sum of all pixels classified as potentially suitable under the binary maps.
276 Second, the northern (i.e., leading or cold) and southern (i.e., trailing or warm) latitudinal limit of the
277 predicted distributional ranges were defined as the 95% and 5% quantile in latitudinal position,
278 respectively, of all pixels classified as potentially suitable. Next, we quantified species thermal
279 response curves for mean annual temperature (BIO1), maximum temperature of the warmest month
280 (BIO5) and minimum temperature of the coldest month (BIO6). Note that we randomly sampled
281 1,000,000 pixels over the potentially suitable area to optimize computational power. For each
282 variable, we derived the cold edge (Q05), the optimum (mode), the warm edge (Q95), and the niche
283 width (Q95 – Q05). Analogous to the model performance, we used BRMs with the same settings to
284 assess differences in model predictions between the SDMs based on the three types of climate data
285 (Table S2). Values of bioclimatic variables were standardized before the analysis to aid model
286 convergence.

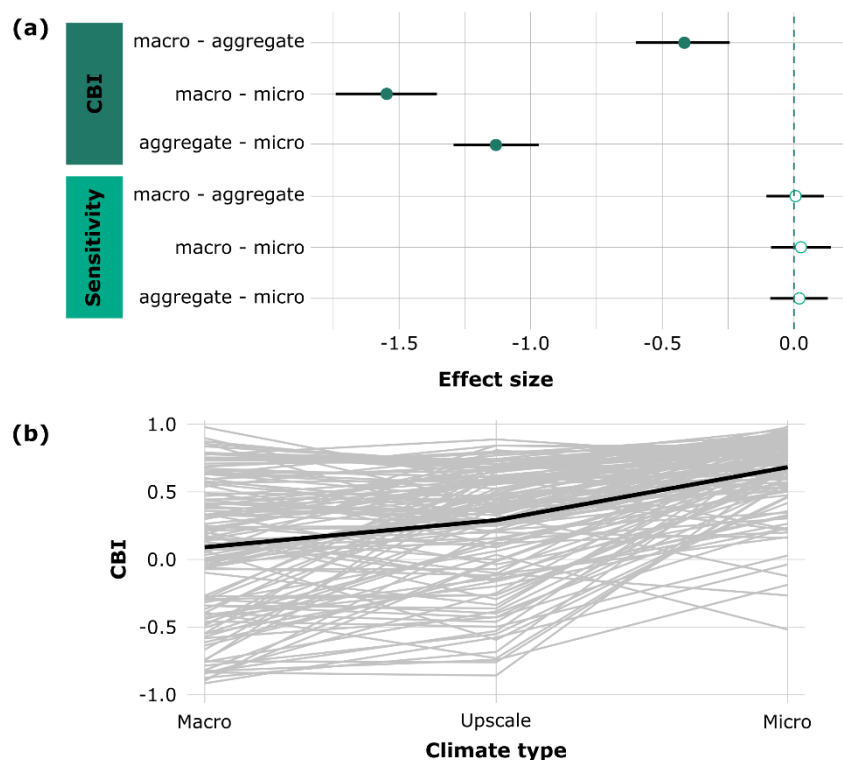
287 Finally, we analyzed whether species are constrained to specific (relative) temperature
288 conditions in their leading and trailing latitudinal limits, as this potentially has important implications
289 for nature conservation. For the trailing and leading edge, we extracted the 5% most southern and
290 northern occurrence records, respectively. Using paired two-sided t-tests ($\alpha = 0.05$), we compared the
291 local temperature conditions of these occurrence points to the surrounding microclimatic conditions
292 over a range of circular buffers (i.e., 100 m, 500 m, 1000 m, 2500 m, 5000 m; Figure S2) around each
293 occurrence record.

294 All calculations were performed in R version 4.1.1 (R Core Team, 2021). The Tier-2 Genius
295 cluster from the high-performance computing facilities of Flanders was used to make the predictions.
296 In order to improve reproducibility, we followed the ODMAP (Overview, Data, Model, Assessment and
297 Prediction) protocol to report on the SDMS in this study (Table S3; Zurell et al., 2020).

298 **RESULTS**

299 **Model performance & sensitivity**

300 We found significant differences ($\alpha = 0.05$) in model performance between models constructed with
 301 (i) macroclimatic (mean CBI = 0.09; se = 0.04) and microclimatic (mean CBI = 0.67; se = 0.02) data, (ii)
 302 macroclimatic and aggregated microclimatic (mean CBI = 0.28; se = 0.04), and (iii) aggregated
 303 microclimatic and microclimatic data (Figure 2). Furthermore, there were no significant differences
 304 between any of the groups regarding the sensitivity of the models. We identified 11 species (i.e.,
 305 *Anemone trifolia*, *Asarum europaeum*, *Clematis recta*, *Cyclamen purpurascens*, *Dictamnus albus*,
 306 *Gagea spathacea*, *Lathyrus vernus*, *Neottia nidus-avis*, *Polystichum aculeatum*, *Ribes spicatum*, and
 307 *Saxifraga hirsuta*) for which the incorporation of microclimatic data did not increase the performance
 308 of the SDMs in comparison to conventional SDM practices, contrarily to the other 129 species for
 309 which microclimate improved performance.

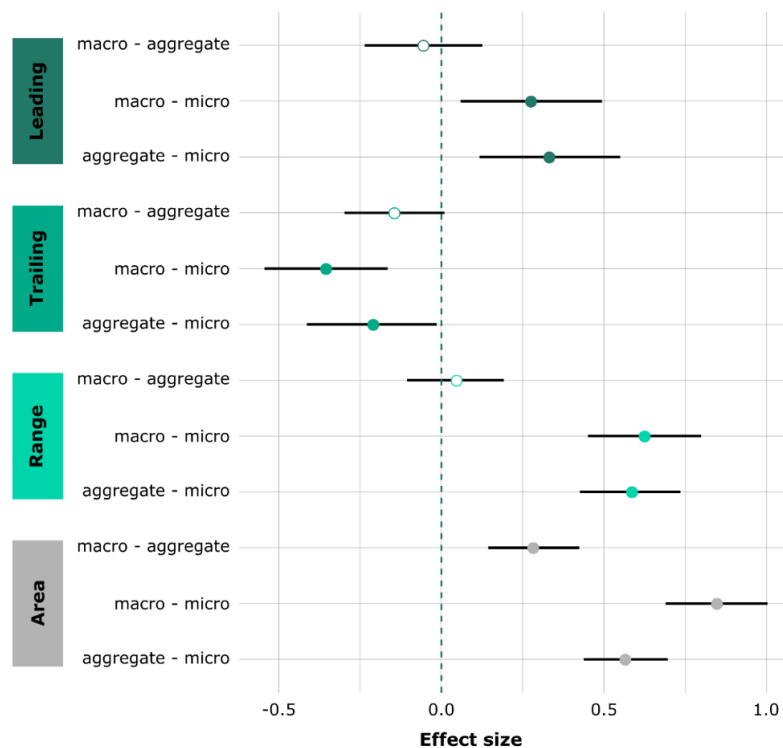


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311 *Figure 2: (a) Pairwise comparison of performance (CBI) and sensitivity between SDMs build with macroclimatic,*
 312 *aggregated microclimatic and microclimatic data. A positive effect size of the comparison reflects a higher model*
 313 *performance and sensitivity in SDMs built with the first group of climate data compared to SDMs built with the*
 314 *second group of climate data. Negative effect sizes reflect the opposite result. Points and associated black error*
 315 *bars correspond to posterior means and 95% highest posterior density intervals of the differences (of the scaled*
 316 *CBI and sensitivity). Significant differences are indicated by full dots whereas non-significant differences are*
 317 *indicated by transparent dots; (b) Parallel coordinate chart indicating the performance of each SDM per species*
 318 *over the three types of climate data (i.e., macroclimatic data, aggregated microclimatic data and microclimatic*
 319 *data). The thick black line shows the average CBI value over each of the three climate types. The GGally package*
 320 *was used to create the parallel coordinate plot (Schloerke et al., 2022).*

321 **Potential suitable area & latitudinal range**

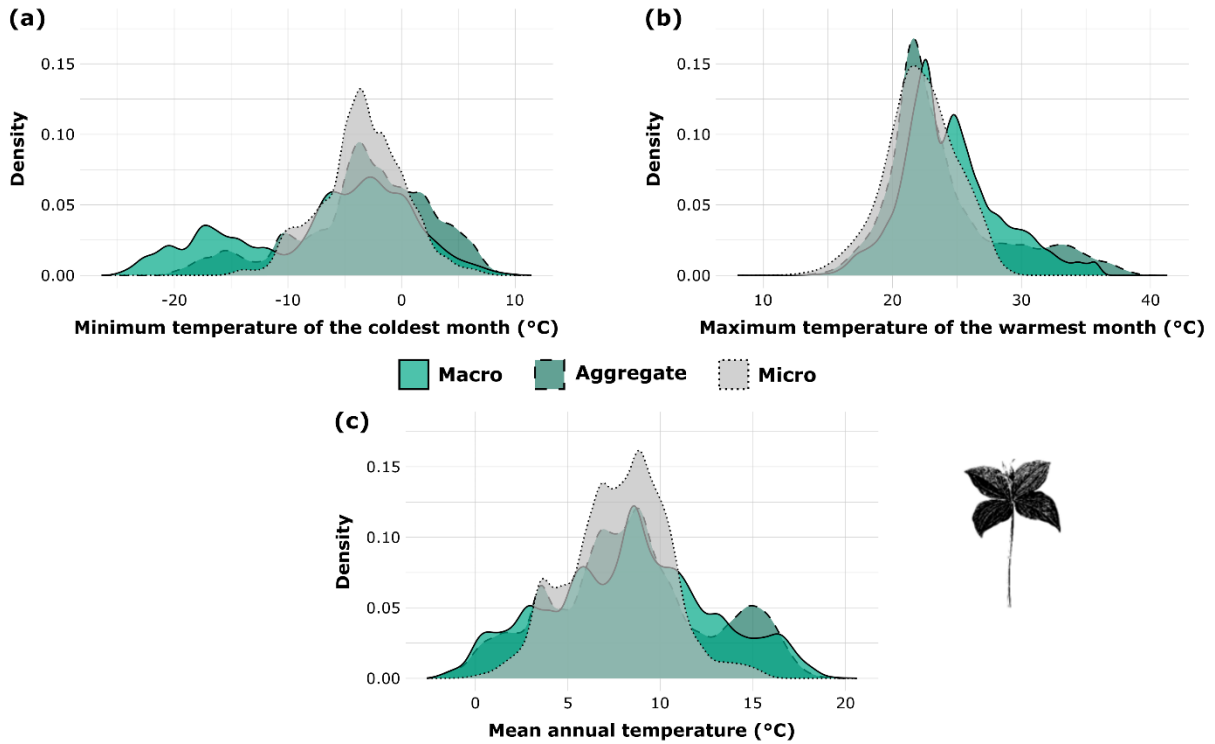
322 A first visual assessment of the binary maps showed clear differences in the potential suitable area
 323 and the potential latitudinal range covered by each species between models calibrated with
 324 macroclimatic data and models calibrated with microclimatic data at the native spatial resolution of
 325 25 m × 25 m (non-aggregated data) (e.g., *Paris quadrifolia*; Figure S3). Indeed, the Bayesian regression
 326 models confirm these visual interpretations (Figure 3). Relative to the native microclimate-based
 327 SDMs, both the leading and trailing edge of the species' distributional ranges are significantly
 328 overestimated when using either macroclimatic or aggregated microclimatic data at 1-km².
 329 Consequently, species' potential latitudinal ranges are significantly smaller when using SDMs
 330 calibrated with microclimatic data (mean = 2,261 km; se = 42 km) in comparison with SDMs calibrated
 331 with aggregated microclimatic data (mean = 2,580 km; se = 43 km) or macroclimatic data (mean =
 332 2,620 km; se = 49 km). Analogous, a species' potential suitable area is significantly smaller when using
 333 SDMs calibrated with microclimatic data (mean = 911,845 km²; se = 30,383 km²) in comparison with
 334 SDMs calibrated with aggregated microclimatic data (mean = 1,148,763 km²; se = 33,527 km²) or
 335 macroclimatic data (mean = 1,268,189 km²; se = 38,274 km²).



336
 337 *Figure 3: Pairwise comparison of the leading edge, trailing edge, latitudinal range, and potential suitable area,*
 338 *respectively between SDMs build with macroclimatic, aggregated microclimatic and microclimatic data. A*
 339 *positive effect size of the comparison reflects more northern leading edges, more northern trailing edges, higher*
 340 *latitudinal ranges and more potentially suitable area in SDMs built with the first group of climate data compared*
 341 *to SDMs built with the second group of climate data. Negative effect sizes reflect the opposite result. Points and*
 342 *associated black error bars correspond to posterior means and 95% highest posterior density intervals of the*
 343 *differences (of the standardized variables). Significant differences are indicated by full dots whereas insignificant*
 344 *differences are indicated by transparent dots.*

345 **Species response curves**

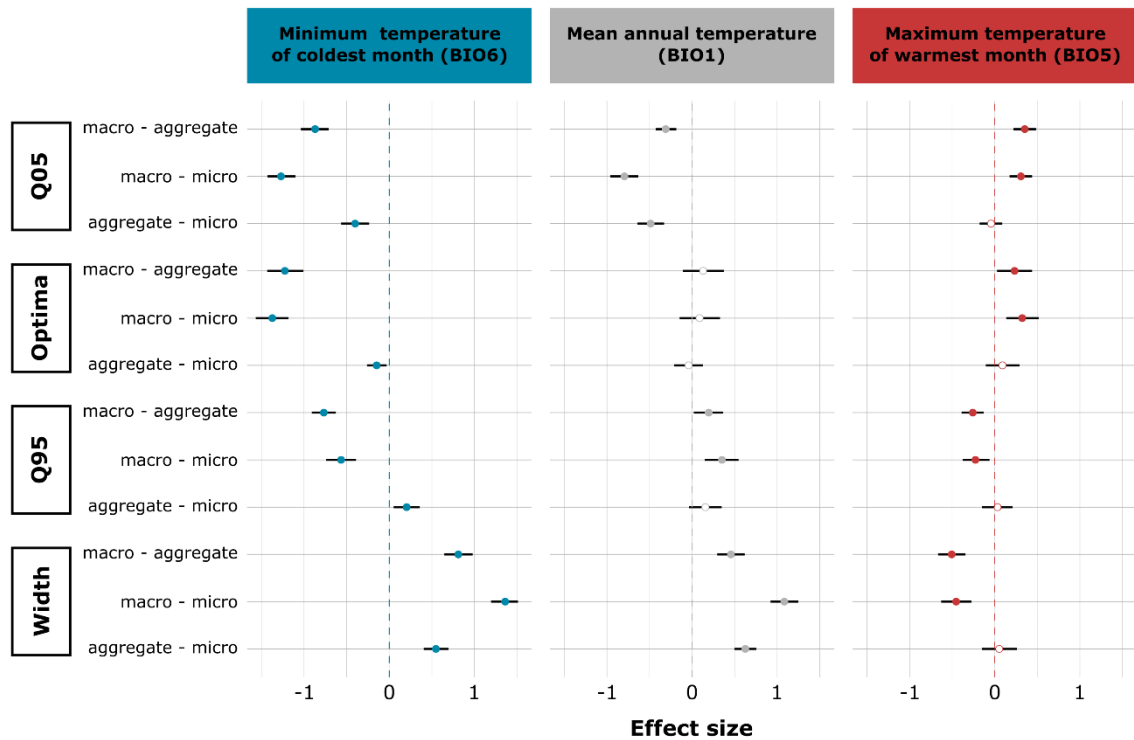
346 A first visual assessment of the response curves showed that microclimate-based response curves of
347 minimum temperature of the coldest month, mean annual temperature and maximum temperature
348 of the warmest month have different optima, and narrower niches compared to macroclimate-based
349 response curves (e.g., *Paris quadrifolia*; Figure 4).



350

351 *Figure 4: Response curves for (a) minimum temperature of the coldest month; (b) maximum temperature of the*
352 *warmest month and (c) mean annual temperature for Paris quadrifolia.*

353 Here, the Bayesian regression models partially supports these visual interpretations (Figure 5). Optima
354 significantly differed between SDMs ran with microclimate and macroclimate data for minimum and
355 maximum temperatures, with warmer optima in minimum temperature and cooler optima in
356 maximum temperature for microclimate-based SDMs relative to macroclimate based SDMs. However,
357 for mean temperature there are no significant differences in optima between the different climate
358 types. Furthermore, the niche width was narrower in minimum and mean temperatures for
359 microclimate-based SDMs relative to macroclimate based SDMs. Surprisingly, the niche width is
360 significantly wider in maximum temperatures for microclimate-based SDMs relative to macroclimate
361 based SDMs.



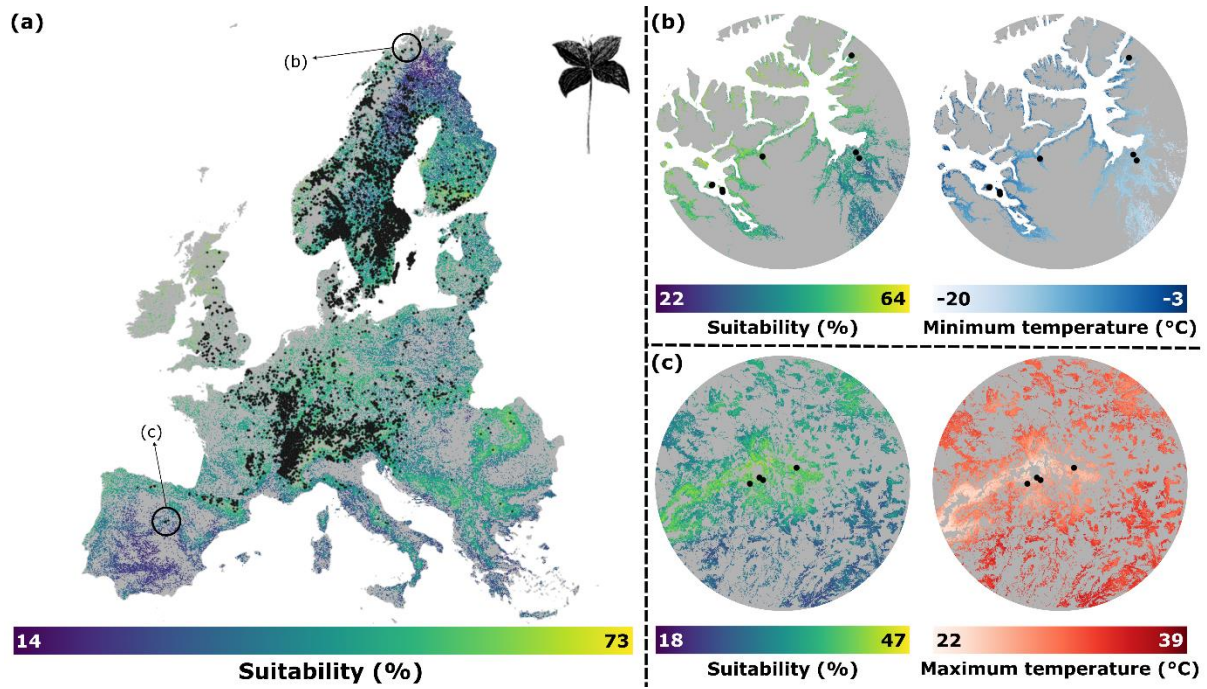
362

363 *Figure 5: Pairwise comparison of the cold edge (Q05), optimum, warm edge (Q95), and niche width, respectively*
 364 *between SDMs built with macroclimatic, aggregated microclimatic and microclimatic data . Each of the*
 365 *comparisons is made for minimum temperature of the coldest month (BIO6), mean annual temperature (BIO1),*
 366 *and maximum temperature of the warmest month (BIO5), respectively. A positive effect size reflects warmer*
 367 *values for the cold edge, optima and warm edge as well as wider niche widths, respectively, in SDMs built with*
 368 *the first group of climate data compared to SDMs built with the second group of climate data. Negative effect*
 369 *sizes reflect the opposite result. Points and associated black error bars correspond to posterior means and 95%*
 370 *highest posterior density intervals of the differences (of the standardized variables). Significant differences are*
 371 *indicated by full dots whereas insignificant differences are indicated by transparent dots.*

372 **Microrefugia**

373 For a buffer of 2500 m, we found that 66% of all the species are constrained in local microrefugia at
 374 their range edges. More specifically, 41% of the species found refuge in warm refugia, relative to the
 375 surrounding landscape, at the leading edge while 49% of the species have remnant populations in cool
 376 refugia, relative to the surrounding landscape, at the trailing edge (e.g., *Paris quadrifolia*; Figure 6).

377



378

379 *Figure 6: (a) Suitability map for Paris quadrifolia resulting from an SDM built with microclimatic data at 25 m ×*
380 *25 m resolution. The black dots represent the occurrence points extracted from GBIF and used as an input to the*
381 *SDMs. We see that the species is located in (b) warm refugia (i.e., higher minimum temperature values in the*
382 *coldest month of the year) at their leading edge and in (c) cool refugia (i.e., lower maximum temperature values*
383 *in the warmest month of the year) at their trailing edge. The grey background shows non-forest areas.*

384 **DISCUSSION**

385 Here, we tested the effect of incorporating three different types of climatic data (i.e., macroclimate,
386 aggregated microclimate matching the spatial resolution of macroclimate and microclimate at its fine
387 spatial resolution) into species distribution models (SDMs) for 140 forest specialist plant species,
388 allowing us to investigate the effect of microclimate as well as the effect of spatial resolution on the
389 performance and predictions of SDMs. We found substantial differences in the model performance
390 (based on the Continuous Boyce Index), indicating that microclimate-based SDMs significantly
391 outperform their conventional (i.e., macroclimate) counterparts and that aggregating microclimate
392 data at coarser spatial resolutions matching macroclimate leads to significant loss in performances.
393 However, the use of aggregated proximal data is still a significant improvement to the use of
394 conventional macroclimate data in SDMs. Furthermore, species response curves derived from the
395 model predictions differed substantially between climate types. These findings highlight the
396 importance of incorporating microclimate data within SDMs, as already postulated by Lembrechts et
397 al. (2018). Certainly within the face of climate change, microclimate-based SDMs are a valuable tool
398 as they allow to identify local refugia for biodiversity conservation. Indeed, up to 66% of the studied
399 species have remnant populations in warm or cold refugia at the leading or trailing edge, respectively.
400 Accordingly, management practices should be in line with the gained insights in order to preserve local
401 microrefugia as they are able to preserve biodiversity despite macroclimatic warming.

402 **The strength of microclimate-based species distribution models**

403 Over the last years, microclimate research focused on improving our understanding of the drivers
404 behind differences between microclimate and macroclimate temperatures (Zellweger et al., 2019) and
405 predicting and mapping microclimate temperatures across space and time (Greiser et al., 2018;
406 Kearney et al., 2019; Lembrechts et al., 2022). Although the drivers behind forest microclimates are
407 relatively well understood, testing how microclimate layers perform within ecological applications
408 such as SDMs has been limited, especially so across large (i.e., continental) spatial extents.

409 Previous research supports our findings on an increased performance of microclimate-based
410 SDMs. For instance, Ashcroft et al. (2008) have already stated that explaining plant species
411 distributions benefits from increasing the accuracy of local temperature, which is also confirmed by
412 Slavich et al. (2014) who have shown that using topoclimate rather than coarse-gridded macroclimatic
413 data leads to improved model performance. On the contrary, Stark & Fridley (2022) did not find
414 significant differences in the performance of microclimate-based and macroclimate-based joint SDMs.
415 Nevertheless, when studying plant species distributions, high-resolution environmental information
416 has overall shown to significantly improve model performance and spatial predictions.

417 The increased performance of microclimate-based SDMs can be explained by several reasons,
418 which are mainly related to the two main input sources of each SDM: the occurrence points and our
419 predictors. First, each occurrence point is subjected to a certain amount of positional error (Wüest et
420 al., 2020). In this study, we only used records with a very low coordinate uncertainty (< 25 m).
421 However, putting such a threshold on the positional error might induce a loss of model power by
422 reducing the sample size of the occurrence points (Guisan et al., 2007). Therefore, many studies often
423 include occurrence records with higher positional uncertainties (e.g., Sanczuk et al., 2022). To deal
424 with these errors within the analysis, it is conventionally suggested to increase the spatial resolution
425 of the analysis to compensate for any positional errors in the occurrence points. However, SDMs are
426 sensitive to changes in the spatial resolution (Chauvier et al., 2022; Manzoor et al., 2018). Decreasing
427 spatial resolution inherently induces a loss of information as the data is smoothed (i.e., aggregated).
428 This comes at the cost of model performance as shown by the CBI values from the models built with
429 aggregated microclimatic data. Therefore, Gábor et al. (2022) strongly recommend to fit models as
430 close as possible to the response grain of the species, meaning that it is recommended to calibrate
431 SDMs with environmental data consistent with the biological scale of the system or organism under
432 study (Randin et al., 2009). For instance, when modelling sessile species (i.e., species with low
433 mobility) or organisms in systems characterized by high environmental heterogeneity, predictors with
434 an increased spatial resolution will be needed to capture the details in their niches more accurately
435 (Araújo et al., 2019; Elith & Leathwick, 2009). Consequently, we expect that the results from this study
436 are not necessarily transferable when studying more mobile species (e.g., birds or mammals) in
437 homogeneous systems (e.g., flat terrain, monoculture plantations). Nevertheless, aggregating
438 proximal data still significantly improves the performance of SDMs relative to the use of conventional
439 macroclimate data in SDMs. Especially when computational capacity is limited, aggregated
440 microclimatic data could be used in order to improve models.

441 **The climate that really matters**

442 The increased availability of microclimatic data products over large spatial extents (Haesen et al.,
443 2023; Lembrechts et al., 2022) opens new avenues within ecological research. With these data, we
444 are able to unravel the hidden niches and describe the conditions that actually matter for species living
445 close to the ground surface (e.g., tree seedlings and forest floor herbs) at very fine spatial resolutions.
446 This study makes clear that species environmental niches derived from conventional macroclimatic
447 data are much wider than one would expect given the buffering effect of forests (De Frenne et al.,
448 2019). Furthermore, the niche estimations based on microclimatic data make it possible to more
449 accurately pinpoint where species could come under pressure due to climate change, facilitating
450 alignment of appropriate management actions (e.g., assisted migration). Indeed, an accurate

451 assessment of species distributional ranges will become vital as many forest specialist species are
452 characterized by slow dispersal rates, up to several meters per year (Hermy et al., 1999; Svenning et
453 al., 2008). It is very unlikely that these species will be able to follow contemporary macroclimate
454 warming, where climate zones are shifting several kilometres each year (Burrows et al., 2011). In this
455 respect, microclimate data and microclimate-based SDMs offer a solution as they will allow us to more
456 accurately assess the velocity of microclimate warming experienced by organisms and its effect on
457 species redistributions. Although outside the scope of this research, this could potentially reveal that
458 the actual velocity of species redistributions does not have to be as high as predicted based on
459 macroclimatic data.

460 When taking a closer look at the trailing and leading edge, we found that 66% of the studied
461 species already persist in cool or warm refugia, respectively (Figure 6; Figure S2). Current SDM
462 practices are unable to identify these microrefugia as conventional macroclimate data represents the
463 overarching free-air temperatures rather than the local temperatures inside these microrefugia
464 (Lenoir et al., 2017), which act at high-spatial resolution. Microclimate-based SDMs thus enable the
465 species-level identification of these microrefugia (Michalak et al., 2020). These insights will thus be
466 crucial for conservation practices as the importance of microrefugia regarding the accumulation and
467 conservation of biodiversity has been widely discussed in the recent scientific literature (Finocchiaro
468 et al., 2022; Nadeau et al., 2022). Indeed, microclimate-based SDMs are a valuable tool to identify
469 areas within the landscape where forest management practices can be aligned (i) to increase the
470 capacity of species and communities to resist climate change (i.e., resistance strategy) or (ii) to
471 facilitate the transformation of communities to species that are well adapted to the novel
472 environmental conditions (Hylander et al., 2022).

473 We would like to note that the goal of this study was to compare different types of climate
474 data for use in SDMs, meaning that we did not necessarily build the best possible model for each
475 species. Therefore, the environmental niches and distributional ranges provided in this study should
476 not be used to undertake conservation actions.

477 **Future improvements**

478 As gridded microclimatic data at 25 m × 25 m is currently only available for European forests, this
479 multi-species study is limited to 140 forest specialist plant species. Here, we do not cover herbaceous
480 plant species living in open habitats such as grasslands or heathlands, as relevant microclimate data
481 at the relevant spatial resolution is not yet available for these habitats. This is mainly due to the fact
482 that measurements from available microclimate sensors are strongly affected by incoming solar
483 radiation when installed in open ecosystems (Maclean et al., 2021), which prevents the development
484 of analogous microclimatic data for these systems. In order to assess whether the results from this

485 study are transferable to other species groups, accurate microclimate data over large spatial extents
486 in open systems is thus urgently needed. Furthermore, additional variables play an important role in
487 plant species distributions but are not yet available at high spatial resolution over large spatial extents.
488 For instance, soil moisture is known to be vital for plant survival, but up till now the topographic
489 wetness index has often been used as a proxy (Kopecký et al., 2021) as high-resolution soil moisture
490 products are hard to obtain. Finally, microclimatic data predicted under future shared socioeconomic
491 pathways (SSPs) will improve insights for future conservation efforts. For instance, knowledge on
492 species reshuffling under climate change or thermophilization of species communities could benefit
493 from such microclimate change predictions. However, forests are very dynamic systems and their
494 structural characteristics – known to influence the forest microclimate – cannot be assumed to remain
495 static over time, which hampers the development of such dynamic products up till now (De
496 Lombaerde et al., 2022). Especially within a warming world, disturbances affecting forest canopies
497 (e.g., drought, pests, storms) will become more frequent and pronounced (De Frenne et al., 2021;
498 Seidl et al., 2017).

499 **CONCLUSIONS**

500 In this study, we performed a comparative analysis between species distribution models (SDMs) with
501 three different sets of climate data: (i) conventional macroclimatic data; (ii) aggregated microclimatic
502 data matching the macroclimatic data in spatial resolution; and (iii) microclimatic data at the native
503 fine spatial resolution that is relevant to the size of the studied organisms (i.e., understory forest
504 specialists here). We conclude that the performance of SDMs for forest specialist species can be
505 significantly improved by incorporating microclimatic data, although this might not necessarily be
506 transferable to other species groups. We would like to emphasize the ability of microclimate-based
507 SDMs to uncover the hidden niche of forest specialist plant species, which has implications for the
508 predicted tolerance of these species at their warm and cold edge. Furthermore, this study makes clear
509 that macroclimatic data is unable to identify warm and cold refugia beyond the range edges of species'
510 distributions. Therefore, we conclude that microclimate-based SDMs are a crucial tool to gain peculiar
511 insights regarding biodiversity conservation within the face of climate change, which is needed to align
512 management actions and prioritize conservation efforts.

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771

772 **Supplementary material**

773 *Table S1: List of the study species, selected from the European forest vascular plant species list (Heinken et al.,*
 774 *2022). We report the amount of occurrence records used within the SDMs after data cleaning.*

Species	Nr. of records	Species	Nr. of records
<i>Aconitum lycoctonum</i> subsp. <i>lasiostomum</i>	0	<i>Hymenophyllum tunbrigense</i>	24
<i>Actaea erythrocarpa</i>	18	<i>Hypericum androsaemum</i>	2595
<i>Actaea spicata</i>	9158	<i>Hypopitys monotropa</i>	3921
<i>Adoxa moschatellina</i>	2365	<i>Impatiens noli-tangere</i>	4548
<i>Aegonychon purpureocaeruleum</i>	422	<i>Impatiens parviflora</i>	12917
<i>Allium ursinum</i>	8970	<i>Inula helvetica</i>	149
<i>Androsace chaixii</i>	14	<i>Knautia drymeia</i>	117
<i>Anemone ranunculoides</i>	2818	<i>Lamium galeobdolon</i>	0
<i>Anemone trifolia</i>	154	<i>Lamium galeobdolon</i>	22345
<i>Arabis turrata</i>	558	<i>Laser trilobum</i>	43
<i>Aremonia agrimonoides</i>	22	<i>Laserpitium nestleri</i>	30
<i>Arenaria procera</i>	0	<i>Lathraea squamaria</i>	3812
<i>Arum italicum</i>	8219	<i>Lathyrus cirrhosus</i>	1
<i>Arum maculatum</i>	7331	<i>Lathyrus niger</i>	1653
<i>Asarum europaeum</i>	1892	<i>Lathyrus venetus</i>	55
<i>Asperula taurina</i>	69	<i>Lathyrus vernus</i>	8234
<i>Brachypodium sylvaticum</i>	12233	<i>Limodorum abortivum</i>	888
<i>Bromopsis benekenii</i>	1632	<i>Lonicera alpigena</i>	698
<i>Bromopsis ramosa</i>	1094	<i>Lonicera nigra</i>	1241
<i>Calamagrostis chalybaea</i>	378	<i>Lunaria rediviva</i>	1365
<i>Calypso bulbosa</i>	7	<i>Luzula forsteri</i>	1103
<i>Campanula latifolia</i>	1393	<i>Luzula luzulina</i>	84
<i>Cardamine bulbifera</i>	4186	<i>Luzula nivea</i>	1032
<i>Cardamine enneaphyllos</i>	392	<i>Luzula pedemontana</i>	19
<i>Cardamine glanduligera</i>	65	<i>Maianthemum bifolium</i>	14196
<i>Cardamine heptaphylla</i>	1768	<i>Melampyrum bohemicum</i>	23
<i>Cardamine pentaphyllos</i>	664	<i>Melica picta</i>	5
<i>Cardamine trifolia</i>	244	<i>Melica uniflora</i>	8231
<i>Carex alba</i>	615	<i>Melittis melissophyllum</i>	4174
<i>Carex digitata</i>	10190	<i>Mercurialis ovata</i>	28
<i>Carex disperma</i>	304	<i>Milium effusum</i>	11644
<i>Carex elongata</i>	2403	<i>Moehringia muscosa</i>	648
<i>Carex fritschii</i>	0	<i>Moneses uniflora</i>	3624
<i>Carex loliacea</i>	562	<i>Neottia cordata</i>	5301
<i>Carex pendula</i>	3397	<i>Neottia nidus-avis</i>	4685
<i>Carex pilosa</i>	150	<i>Neottianthe cucullata</i>	0
<i>Carex remota</i>	7905	<i>Omphalodes scorpioides</i>	16
<i>Carex strigosa</i>	367	<i>Onoclea struthiopteris</i>	0
<i>Cephalanthera damasonium</i>	1508	<i>Orchis spitzelii</i>	6
<i>Cerastium sylvaticum</i>	4	<i>Orobanche hederarum</i>	384
<i>Chimaphila umbellata</i>	1514	<i>Orobanche lucorum</i>	5
<i>Chrysosplenium oppositifolium</i>	2490	<i>Orobanche salviae</i>	14

<i>Cinna latifolia</i>	898
<i>Circaea alpina</i>	1744
<i>Circaea lutetiana</i>	9695
<i>Circaea x intermedia</i>	275
<i>Cirsium carniolicum</i>	8
<i>Clematis recta</i>	102
<i>Clinopodium menthifolium</i>	486
<i>Coptidium lapponicum</i>	1156
<i>Coronilla coronata</i>	45
<i>Corydalis cava</i>	2087
<i>Cyclamen hederifolium</i>	1805
<i>Cyclamen purpurascens</i>	752
<i>Cypripedium calceolus</i>	710
<i>Cystopteris sudetica</i>	19
<i>Dactylis glomerata</i> subsp. <i>lobata</i>	999
<i>Daphne laureola</i>	4501
<i>Daphne mezereum</i>	12362
<i>Dictamnus albus</i>	524
<i>Digitalis purpurea</i>	22045
<i>Diplazium sibiricum</i>	96
<i>Drymochloa drymeja</i>	8
<i>Drymochloa sylvatica</i>	2544
<i>Dryopteris remota</i>	13
<i>Epipactis albensis</i>	0
<i>Epipactis bugacensis</i>	1
<i>Epipactis fageticola</i>	0
<i>Epipactis greuteri</i>	0
<i>Epipactis leptochila</i>	29
<i>Epipactis microphylla</i>	69
<i>Epipactis nordeniorum</i>	0
<i>Epipactis phyllanthes</i>	34
<i>Epipactis placentina</i>	0
<i>Epipactis pontica</i>	0
<i>Epipactis purpurata</i>	210
<i>Epipactis tallosii</i>	0
<i>Epipogium aphyllum</i>	144
<i>Euonymus latifolius</i>	245
<i>Euphorbia amygdaloides</i>	7039
<i>Festuca flavescens</i>	8
<i>Gagea spathacea</i>	263
<i>Galium aristatum</i>	45
<i>Galium intermedium</i>	3
<i>Galium laevigatum</i>	19
<i>Galium odoratum</i>	18818
<i>Galium rotundifolium</i>	1155
<i>Galium sylvaticum</i>	590
<i>Galium triflorum</i>	391

<i>Orthilia secunda</i>	7210
<i>Oxalis acetosella</i>	27704
<i>Paeonia mascula</i>	29
<i>Paris quadrifolia</i>	15002
<i>Pilosella hybrida</i>	0
<i>Poa remota</i>	778
<i>Poa stiriaca</i>	0
<i>Polygonatum multiflorum</i>	7309
<i>Polystichum aculeatum</i>	1935
<i>Polystichum braunii</i>	561
<i>Polystichum setiferum</i>	1662
<i>Prenanthes purpurea</i>	2892
<i>Prunus lusitanica</i>	322
<i>Pulmonaria collina</i>	0
<i>Pulmonaria longifolia</i>	573
<i>Pulmonaria obscura</i>	2081
<i>Pulmonaria officinalis</i>	1965
<i>Pulmonaria saccharata</i>	103
<i>Pyrola chlorantha</i>	3752
<i>Pyrola media</i>	1304
<i>Pyrola minor</i>	3735
<i>Ranunculus cassubicus</i>	187
<i>Rhamnus alpina</i>	313
<i>Ribes spicatum</i>	1528
<i>Rumex sanguineus</i>	3069
<i>Ruscus aculeatus</i>	10151
<i>Sanicula europaea</i>	5624
<i>Saxifraga geranioides</i>	14
<i>Saxifraga hirsuta</i>	211
<i>Saxifraga umbrosa</i>	92
<i>Schedonorus giganteus</i>	2742
<i>Scilla lilio-hyacinthus</i>	126
<i>Scrophularia alpestris</i>	115
<i>Scrophularia peregrina</i>	38
<i>Scutellaria altissima</i>	73
<i>Sedum cepaea</i>	41
<i>Senecio hercynicus</i>	169
<i>Senecio nemorensis</i> subsp. <i>jacquinianus</i>	8
<i>Silene viridiflora</i>	10
<i>Soldanella montana</i>	42
<i>Spiraea japonica</i>	234
<i>Stachys sylvatica</i>	13096
<i>Staphylea pinnata</i>	243
<i>Stellaria longifolia</i>	795
<i>Stellaria nemorum</i>	587
<i>Symphytum cordatum</i>	4
<i>Trifolium rubens</i>	254

Geranium lanuginosum	56
Geum sylvaticum	181
Goodyera repens	21442
Gymnocarpium dryopteris	9594
Hacquetia epipactis	103
Helleborus niger	957
Helleborus viridis	1162
Hepatica nobilis	28312
Hesperis sylvestris	3
Hordelymus europaeus	2768

Trochiscanthes nodiflora	14
Valeriana pyrenaica	56
Veronica montana	2714
Veronica urticifolia	892
Vicia dumetorum	136
Vicia pisiformis	126
Viola jordanii	1
Viola mirabilis	2449
Viola pseudomirabilis	0
Viola reichenbachiana	6726

775

776 Table S2: Final model structures of the Bayesian regression models indicating which transformations,
777 distributions, priors and formulas were used (modeled with brms package). Phi, sigma and alpha are
778 distributional parameters representing the precision (beta distribution), standard deviation (gaussian and
779 skewed normal distribution) and skewness (skewed normal distribution) respectively. The adapt_delta argument
780 is used to adapt sampling speeds (default value = 0.8), with higher values corresponding to slower sampling
781 speeds and more robust to posterior distributions. Q05, optimum and Q95 refer to cold edge, optimum and warm
782 edge of the bioclimatic variables.

RESPONSE		Transformation	Distribution	PRIORS	Formula
CBI		rescaled	beta	standard	CBI ~ type + (1 species) phi ~ type
SENSITIVITY		/	beta	standard	sensitivity ~ type + (1 species) phi ~ type
AREA		standardized	skewed normal	user-defined*	area ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9
EDGES	Leading	standardized	skewed normal	user-defined*	leading ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9
	Trailing	standardized	skewed normal	standard	trailing ~ type + (1 species) sigma ~ type alpha ~ type
	Range	standardized	skewed normal	user-defined*	range ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9
	Q05	standardized	skewed normal	standard	BIO1_Q05 ~ type + (1 species) sigma ~ type alpha ~ type
BIO1	Optimum	standardized	skewed normal	standard	BIO1_opt ~ type + (1 species) sigma ~ type alpha ~ type
	Q95	standardized	skewed normal	standard	BIO1_Q95 ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9
	Width	standardized	skewed normal	standard	BIO1_width ~ type + (1 species) sigma ~ type alpha ~ type
BIO5	Q05	standardized	gaussian	standard	BIO5_Q05 ~ type + (1 species)
	Optimum	standardized	skewed normal	standard	BIO5_opt ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.99
	Q95	standardized	skewed normal	standard	BIO5_Q95 ~ type + (1 species) sigma ~ type alpha ~ type
	Width	standardized	skewed normal	user-defined*	BIO5_width ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9
BIO6	Q05	standardized	skewed normal	user-defined*	BIO6_Q05 ~ type + (1 species)

					<pre>sigma ~ type alpha ~ type adapt_delta = 0.99</pre>
	Optimum	standardized	skewed normal	standard	<pre>BIO5_opt ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9</pre>
	Q95	standardized	skewed normal	standard	<pre>BIO6_Q95 ~ type + (1 species) sigma ~ type alpha ~ type</pre>
	Width	standardized	skewed normal	standard	<pre>BIO6_width ~ type + (1 species) sigma ~ type alpha ~ type adapt_delta = 0.9</pre>

783 *User-defined priors: c(set_prior("normal(0,3)", class = "b"), set_prior("normal(0,5)", class = "b", par="alpha"), set_prior("student_t(3, 0,
784 2.5)", class = "b", dpar="sigma"))

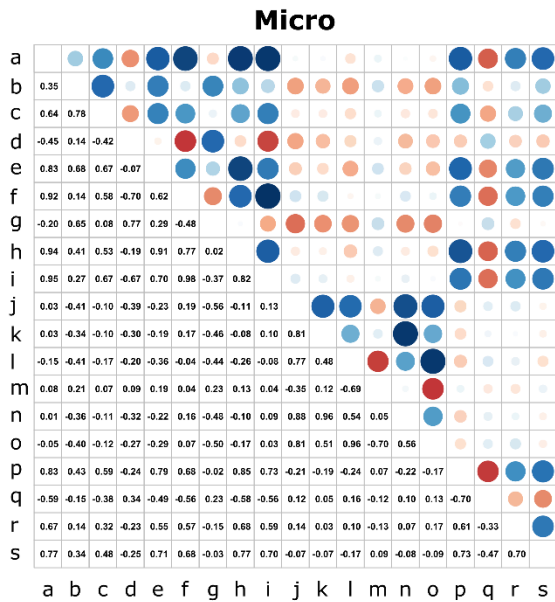
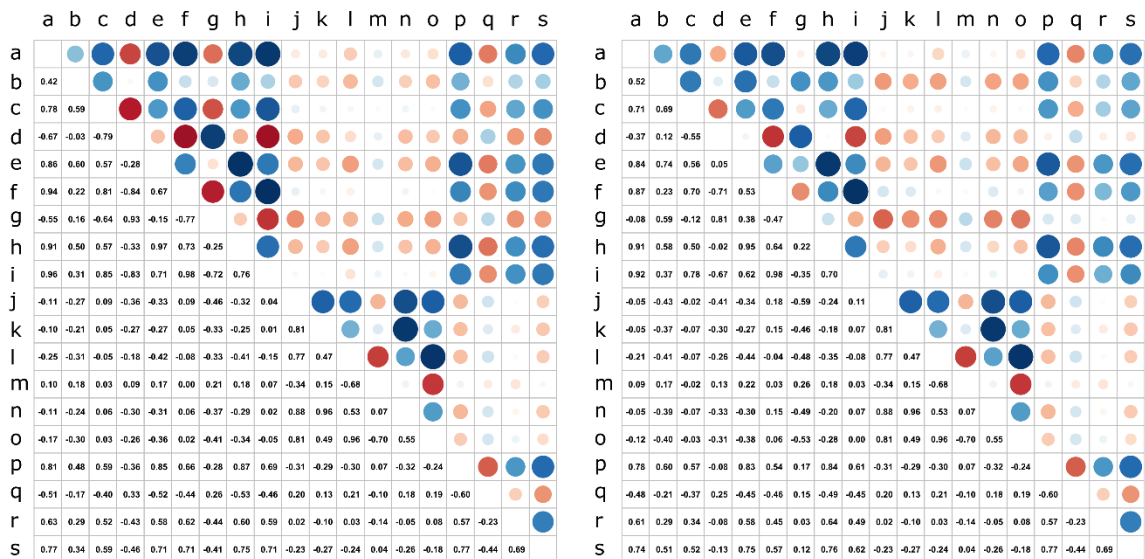
OVERVIEW	
<i>Authorship</i>	<ul style="list-style-type: none"> • Authors: Stef Haesen, Jonathan Lenoir, Eva Gril, Pieter De Frenne, Jonas J. Lembrechts, Martin Kopecký, Martin Macek, Matěj Man, Jan Wild, Koenraad Van Meerbeek • Contact email: stef.haesen@kuleuven.be • Title: Uncovering the hidden niche: incorporating microclimate temperature into species distribution models • DOI: Not applicable
<i>Model objective</i>	<ul style="list-style-type: none"> • Objective: Mapping / interpolation • Target outputs: logistic maps (i.e, continuous habitat suitability index) & binary maps (i.e., suitable vs. unsuitable habitat)
<i>Taxon</i>	forest plant specialist species
<i>Location</i>	Europe
<i>Scale of analysis</i>	<ul style="list-style-type: none"> • Spatial extent (Longitude / Latitude): 8°E - 50°W ; 33°N – 67°N • Spatial resolution: 25 m × 25 m (microclimatic data) & 1 km × 1 km (macroclimatic data & aggregated microclimatic data) • Temporal extent / time period: 2000 - 2020 • Type of extent boundary: rectangular
<i>Biodiversity data</i>	<ul style="list-style-type: none"> • Observation type: human observations (GBIF) • Response / Data type: presence-only
<i>Type of predictors</i>	climatic, edaphic
<i>Conceptual model</i>	<ul style="list-style-type: none"> • Hypotheses about species-environment relationships: As forests are known to buffer temperatures, we hypothesize that forest specialist plant species respond to warmer minimum temperatures and lower maximum temperatures as perceived by the free-air temperature data.
<i>Assumptions</i>	<p>We assumed that:</p> <ul style="list-style-type: none"> ○ Species are at (pseudo-) equilibrium with their environment. ○ The offset values from the long-term average period of 1970-2000 are still valid for the period 2000-2020. These are used to spatially-downscale the 4 km × 4 km TerraClimate data to 1 km × 1 km.
<i>SDM algorithms</i>	<ul style="list-style-type: none"> • Algorithms: We fitted MaxEnt models, which were chosen due to the presence-only character of the occurrence records. • Model complexity: MaxEnt models were built with linear, quadratic and product features. Regularization multipliers ranges from 0.5 to 5. • Ensembles: Not applicable
<i>Model workflow</i>	Only weakly correlated predictors were retained in the analysis. We

	performed parameter tuning for the features and regularization multipliers in MaxEnt models based on AICc, using the ENMeval 2.0 package. Model performance was assessed using the Continuous Boyce Index (CBI)
<i>Software</i>	<ul style="list-style-type: none"> • Software: Analyses were conducted in R version 4.1.1 and models were constructed using ENMeval v2.0.0 with maxnet package v0.1.4. • Data availability: The raw biodiversity data is available through https://doi.org/10.15468/dl.kf533a. ForestClim is freely-available through https://doi.org/10.6084/m9.figshare.22059125.
DATA	
<i>Biodiversity data</i>	<ul style="list-style-type: none"> • Taxon names: All species are listed in Table S1 • Ecological level: individual point data • Data source: GBIF (https://doi.org/10.15468/dl.kf533a) • Sampling design: random • Sample size: Amount of records per species are listed in Table S1 • Regional mask: We clipped the data to the boundary of the study area • Data cleaning / filtering: The occurrence data were filtered in the following sequential steps: (1) only records of ‘human observations’ were selected; (2) records with an unknown coordinate uncertainty or coordinate uncertainty larger than 25 m were excluded; (3) records located at country or capital centroids and biodiversity institutions were omitted; (4) duplicate records were removed; (5) only records observed during our climatic reference period (2000-2020) were selected; (6) records were spatially thinned to one random observation per 25 m × 25 m grid cell; and (7) we omitted species with less than 50 cleaned occurrence records. • Background data: Background data were generated by sampling an equal amount of background points as occurrence points based on a 2D kernel-density estimate of the occurrence points. • Errors and biases: We performed spatial thinning of occurrence points to account for sampling bias. Furthermore, background data was sampled according to a 2D-kernel density function, which introduces an equal amount of sampling bias within the background data as in the presence-only data. We only used records with a very low coordinate uncertainty (< 25 m) in order to minimize positional error on the occurrence points.
<i>Data partitioning</i>	We allocated 80% of the occurrence points to a spatial block cross-validation procedure, whereas 20% is kept for independent evaluation.
<i>Predictor variables</i>	<ul style="list-style-type: none"> • Predictor variables: <ul style="list-style-type: none"> ○ Climate: maximum temperature of the warmest month

	<p>(BIO5), minimum temperature of the coldest month (BIO6), mean annual precipitation (BIO12), and precipitation seasonality (BIO15).</p> <ul style="list-style-type: none"> ○ Edaphic variables: cation exchange capacity and soil clay content ● Data sources: Macroclimatic data was collected from the TerraClimate database, whereas microclimatic data was downloaded from ForestClim. SoilGrids was used for the edaphic variables. ● Spatial resolution: 25 m × 25 m (microclimatic data) & 1 km × 1 km (macroclimatic data & aggregated microclimatic data) ● Extent: 8°E - 50°W ; 33°N – 67°N ● Geographic projection: ETRS89/LAEA ● Time period: 2000-2020 ● Data processing: Bilinear interpolation was used in case data needed to be spatially-downscaled, whereas data aggregation (i.e., averaging) was used when data needed to be spatially-upscaled. All layers were reprojected to ETRS89/LAEA, if needed.
MODEL	
<i>Variables preselection</i>	We started from the conventional set of nineteen bioclimatic variables. Based on Booth (2022), we excluded bioclimatic variables combining both temperature and precipitation data (i.e., BIO8 = mean temperature of the wettest quarter; BIO9 = mean temperature of the driest quarter; BIO18 = precipitation of the warmest quarter; BIO19 = precipitation of the coldest quarter)
<i>Multicollinearity</i>	Highly correlated variables (Spearman correlation coefficients > 0.7) were removed from the analysis in order to reach the most parsimonious model. When excluding one of the correlated covariate pair, we retained variables which are known to be more important for plant species distribution and which are important for our further analyses (e.g., BIO5 & BIO6)
<i>Model settings</i>	MaxEnt models were built with linear, quadratic and product features. Regularization multipliers ranges from 0.5 to 5
<i>Non-independence</i>	We accounted for spatial autocorrelation by implementing a spatial block-cross validation.
<i>Threshold selection</i>	We used a 10% training presence as a threshold, meaning that the suitable area contains 90% of the original occurrence records
ASSESSMENT	
<i>Performance statistics</i>	<ul style="list-style-type: none"> ● Performance statistics estimated on validation data: The Akaike Information Criterion for small sample sizes (AICc) was used to select the best candidate models ● Performance statistics estimated on testing data: Model

	performance was assessed using the Continuous Boyce Index (CBI)
<i>Plausibility checks</i>	Maps of modelled predictions were checked by experts for an ad-hoc subset of species.
PREDICTION	
<i>Prediction output</i>	<ul style="list-style-type: none"> • Prediction unit: logistic maps (i.e, continuous habitat suitability index) & binary maps (i.e., suitable vs. unsuitable habitat) • Post-processing: Non-forested areas where masked out

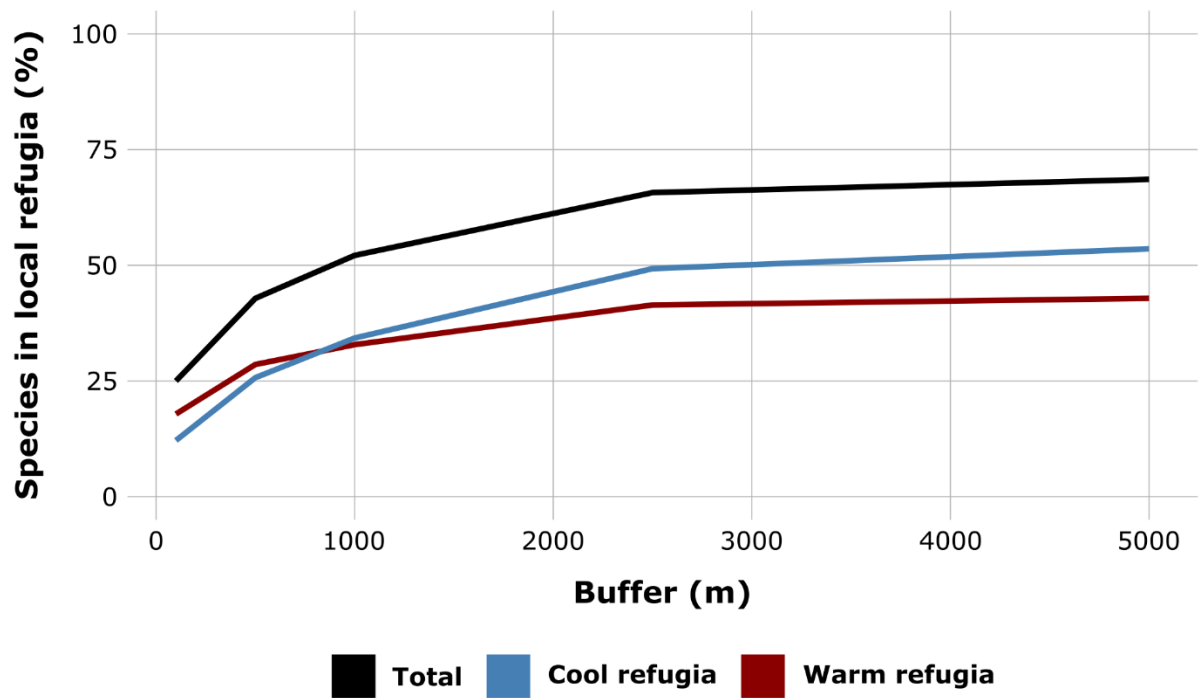
786



- a - BIO1 - mean annual temperature
- b - BIO2 - mean diurnal range
- c - BIO3 - isothermality
- d - BIO4 - temperature seasonality
- e - BIO5 - maximum temperature of the warmest month
- f - BIO6 - minimum temperature of the coldest month
- g - BIO7 - temperature annual range
- h - BIO10 - mean temperature of the warmest quarter
- i - BIO11 - mean temperature of the coldest quarter
- j - BIO12 - annual precipitation
- k - BIO13 - precipitation of the wettest month
- l - BIO14 - precipitation of the driest month
- m - BIO15 - precipitation seasonality
- n - BIO16 - precipitation of the wettest quarter
- o - BIO17 - precipitation of the driest quarter
- p - bulk density
- q - cation exchange capacity
- r - clay content
- s - pH

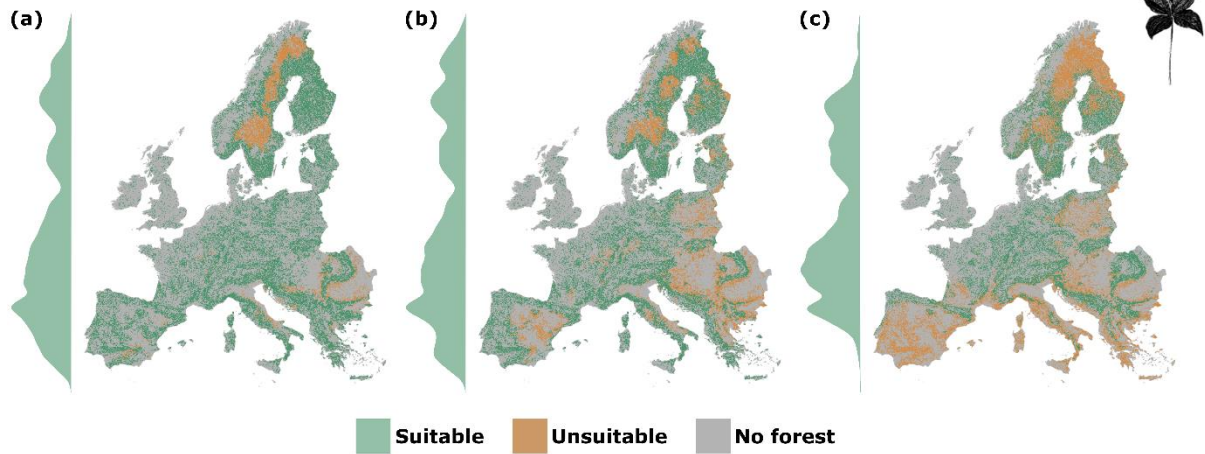
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788 *Figure S1: Correlation matrix containing all Spearman correlation coefficients (r^2) between all combinations of*
 789 *quantitative variables. Highly correlated variables (Spearman correlation coefficients > 0.7) were removed from*
 790 *the analysis (Dormann et al., 2013). The final selection of covariates encompassed two temperature variables*
 791 *(maximum temperature of the warmest month (BIO5) and minimum temperature of the coldest month (BIO6)),*
 792 *two precipitation variables (mean annual precipitation, (BIO12) and precipitation seasonality (BIO15)) and two*
 793 *edaphic variables (cation exchange capacity and soil clay content).*



794

795 *Figure S2: Range of circular buffers (i.e., 100 m, 500 m, 1000 m, 2500 m, 5000 m) over which we quantified*
 796 *whether or not a species was located in cool or warm refugia. For the trailing and leading edge, we extracted the*
 797 *5% most southern and northern occurrence records, respectively. Using paired t-tests ($\alpha = 0.05$), we compared*
 798 *the local temperature conditions of these occurrence points to the surrounding microclimatic conditions over*
 799 *these circular buffers around each occurrence record.*



800

801 *Figure S3: Binary maps indicating the potential suitable area for Paris quadrifolia. These maps are the result of*
 802 *species distribution modelled using (a) macroclimate data at a spatial resolution of 1 km × 1 km, (b) aggregated*
 803 *microclimatic data at a spatial resolution of 1 km × 1 km and (c) microclimatic data at the native spatial resolution*
 804 *of 25 m × 25 m over the 2000-2020 period. Density of suitable pixels along the latitudinal gradient is represented*
 805 *at the left side, respectively.*